

## Introduction

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The delivery of foreign DNA into eukaryotic cells is one of the most common molecular biology techniques to study biological mechanisms. However, unlike transformed cell lines, the efficient transfection of primary cells can be a problem. EndoFectagen is a cationic polymer-based transfection system specifically designed and optimized for efficient transfection of primary endothelial cells. Transfection with EndoFectagen can be carried out in the presence of antibiotics and serum. Instead of normal two-day transfection, an optimized one-day transfection procedure can be performed for time-saving and highly reproducible transfection. One ml of EndoFectagen reagent is sufficient for up to 250 transfections per well in 96-well plate.

## Storage/Handling

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Upon receipt, aliquot and store EndoFectagen reagent A at -20°C, avoid repeated freezing/thawing cycles. Once thawed, store EndoFectagen reagent A at 4°C and use in a month. EndoFectagen reagent B can be kept at 4°C.

## Quality Control

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Each lot of EndoFectagen is performance tested by transfecting Human Umbilical Vessel Endothelial Cells (HUVECs, Cat. No. 8000, ScienCell™) with Promega® pSV-bata-Galactosidase control vector. Gene expression is assayed by X-gal staining 24 hours post transfection. Typically, 30-60% transfection efficiency can be achieved (Figure 1).

## Procedures for Transfecting Adherent Cells in 96-well Plate\*

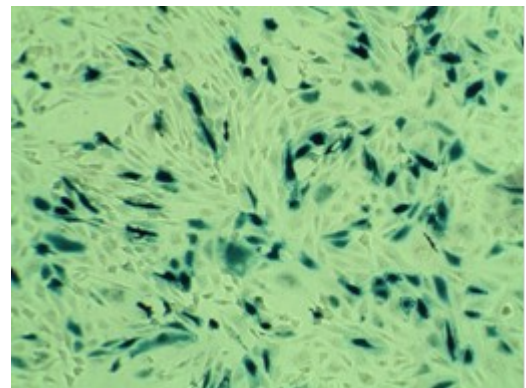
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### A. Preparation of cells

1. On the day of transfection, coat 96-well plate with poly-L-lysine at 2 µg/cm<sup>2</sup>. Incubate at 37°C for 2-4 hours. Rinse the poly-L-lysine coated wells with sterile deionized H<sub>2</sub>O twice before seeding of cells. The pre-coating of poly-L-lysine ensures good and even endothelial cell adhesion.
2. Select a flask of endothelial cells with 60-80% confluency, harvest and dilute cells in Endothelial Culture Medium to give a final concentration of  $\sim 1.1 \times 10^5$  cells/ml.

### B. Transfection complex formation

1. Prepare plasmid DNA in sterile deionized H<sub>2</sub>O to give a final concentration of 1 µg/µl. To achieve successful transfection, high quality DNA with OD<sub>260</sub>/OD<sub>280</sub> of 1.8 or greater is recommended.
2. For each well, add 0.5 µl plasmid DNA, 10.5 µl sterile deionized H<sub>2</sub>O and 5 µl EndoFectagen reagent B into a 1.5 ml sterile plastic tube. Vortex gently and spin down briefly. Then add 4 µl EndoFectagen reagent A to make the total volume of the transfection mixture to be 20 µl, vortex for 5 seconds and spin down. Incubate at room temperature for 20-30 min.



**Figure 1.** HUVECs expressing β-galactosidase after transfection using EndoFectagen.

### C. Incubation of cells with transfection mixture

1. Plate 180  $\mu\text{l}$  of cell suspension ( $\sim 1.1 \times 10^5$  cells/ml) in each well to give  $\sim 2 \times 10^4$  cells per well.
2. Add 20  $\mu\text{l}$  of transfection mixture to each well. Mix by gently rocking the plate side-to-side.
3. Culture the cells for  $\sim 24$  hours under standard conditions. Or perform a medium change after 4-6 hours' incubation with transfection mixtures, replace with 200  $\mu\text{l}$  fresh culture medium, and culture for additional 16-18 hours. Generally longer incubation time with transfection mixture results in increased transfection efficiency and decreased cell viability. For microvascular endothelial cells, relatively short incubation time with transfection mixtures (e.g. 3-4 hours) may be necessary to help cell survive better.
4. Harvest cells 24 hours post transfection and assay for gene expression.

\*The amounts of cells and various transfection reagents mentioned in the instruction are recommended for performing transfection in 96-well plate. For transfection in larger size wells, the amounts of endothelial cells and transfection reagents (DNA, sterile deionized H<sub>2</sub>O and EndoFectagen reagents A&B) should be scaled up according to the surface area of the wells (Table 1).

**Table 1.** Recommended quantities of endothelial cells and EndoFectagen reagents per well.

Culture Vessel	Growth Area (cm <sup>2</sup> /well)	# of cells	1 $\mu\text{g}/\mu\text{l}$ DNA stock ( $\mu\text{l}$ )	Sterile DI H <sub>2</sub> O ( $\mu\text{l}$ )	EndoFectagen reagent B ( $\mu\text{l}$ )	EndoFectagen reagent A ( $\mu\text{l}$ )	ECM ( $\mu\text{l}$ )
96-well plate	0.35	20,000	0.5	10.5	5	4	180
48-well plate	0.8	45,000	1.1	24	11.4	9.1	411
24well plate	2.0	115,000	2.9	60	29	23	1029
12-well plate	4.0	230,000	5.7	120	57	46	2057
6-well plate	9.6	550,000	13.7	288	137	110	4937