

Glutamate Assay (GLU) Cat. No. 8338 100 Tests in 96-well plate

Introduction

Glutamate is a key molecule in cellular metabolism and an important mammalian neurotransmitter that is believed to be involved in a number of neurological diseases such as lateral sclerosis, lathyrism, autism and Alzheimer's disease. Glutamate is also widely used as a flavor enhancer in the food industry. This colorimetric assay is based on glutamate dehydrogenase catalyzed oxidation of glutamate, in which the formed NADH reduces a terazolium salt, MTT. The intensity of the product color, which exhibits maximum absorbance at 570 nm, is proportional to the glutamate concentration in the sample.

Kit Components

| Cat. No. | # of vials | Reagent | Quantity | Storage |
|----------|------------|--------------------|----------|---------|
| 8338a | 1 | Assay buffer | 10 mL | 4°C |
| 8338b | 1 | Glutamate standard | 1 mL | 4°C |
| 8338c | 1 | NAD | 0.4 mL | -20°C |
| 8338d | 1 | Substrate mix | 1.5 mL | -20°C |
| 8338e | 1 | Enzyme | 0.1 mL | -20°C |

Product Use

This kit measures glutamate level of samples from cells, serum, plasma, tissue, and food extracts. It is for research purposes only and not for use in animals, humans, or diagnostic procedures.

Quality Control

Data from Glutamate Assay of glutamate solutions with concentrations ranging from 0.02 to 1 mM show a linear relationship between OD_{570nm} and glutamate concentration (Figure 1).

Shipping and Stability

Shipped on dry ice and the kit is stable for one year when handled properly.

Procedure (96-well plate)

A. Preparation of glutamate standard

1. Add 1 μ L of glutamate standard (8338b) to 99 μ L of assay buffer (8338a) to make a 0.1 mL solution of 1 mM glutamate. Dilute standard as follows.

| No | 1mM(μL) | Assay buffer(μL) | Volume (µL) | Glutamate (mM) |
|----|----------|-------------------|--------------|----------------|
| 1 | 10 | 0 | 10 | 1 mM |
| 2 | 7.5 | 2.5 | 10 | 0.75 mM |
| 3 | 5 | 5 | 10 | 0.5 mM |
| 4 | 2.5 | 7.5 | 10 | 0.25 mM |
| 5 | 1 | 9 | 10 | 0.1 mM |
| 6 | 0.5 | 9.5 | 10 | 0.05 mM |
| 7 | 0 | 10 | 10 | 0 mM |

2. Transfer 10 μL each glutamate standard into wells of the 96-well flat bottom plate.

B. Preparation of test samples

- 1. Cells or tissues can be homogenized in 4 volumes of the assay buffer (8338a). Centrifuge the samples at $13,000 \times g$ for 10 minutes to remove insoluble material. The soluble fraction may be assayed directly.
- 2. Endogenous NADH from cell or tissue extracts generates background in the glutamate assay. To remove the NADH background, the same amount of sample as the control can be tested in the absence of enzyme (8338e).
- 3. Samples should be serial diluted to make sure the readings are within the standard curve range. Prepare test samples to a final volume of $10 \mu L/well$ on the 96-well flat bottom plate.

C. Working reagent preparation and measurements

- 1. For each well of reaction, prepare working reagent by mixing 70 μL assay buffer (8338a), 4 μL NAD (8338c), 15 μL substrate mix (8338d), and 1 μL enzyme (8338e). When a control is required to remove background from a sample, prepare a control working reagent by mixing 71 μL assay buffer (8338a), 4 μL NAD (8338c) and 15 μL substrate mix (8338d).
- 2. Add 90 μL of working reagent mix into each well of the 96-well plate containing glutamate standard, samples, and blank. Add 90 μL of control working reagent mix into each well of the 96-well plate containing samples for control. Incubate for 20 minutes at room temperature protected from light.
- 3. Read the absorbance at 570 nm with an ELISA plate reader.

D. Calculations

- 1. Subtract the OD_{570nm} value of the blank from the OD_{570nm} values obtained with all other standard and samples to get ΔOD_{570nm} value.
- 2. Based on the calibrated ΔOD_{570nm} of the glutamate standard, make a standard curve by plotting ΔOD_{570nm} as a function of glutamate concentration (See Figure 1 for a typical standard curve). Determine the equation and R^2 value of the trend line.
- 3. For samples requiring control without enzyme, subtract the ΔOD_{570nm} without enzyme value from the ΔOD_{570nm} with enzyme value and use this $\Delta \Delta OD_{570nm}$ value to determine the sample glutamate concentration from the standard curve.
- 4. Because the equation of the trend line of the standard curve is y = Ax + B, calculate the glutamate concentration of test samples as follows:

[glutamate] =
$$\frac{\Delta\Delta OD_{570nm}}{A}$$

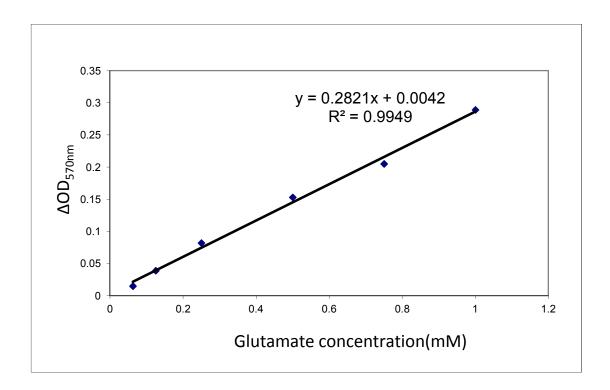


Figure 1. A typical glutamate standard curve measured by ScienCell™ Glutamate Assay kit