Human Gallbladder Fibroblasts (HGBF)

Catalog Number: 5430

Cell Specification:

Fibroblasts are mesenchymal cells derived from the embryonic mesoderm. Because they are one of the easiest types of cells to grow in culture, they have been extensively used for a wide range of cellular and molecular studies. Additionally, their durability makes them amenable to a wide variety of manipulations ranging from studies employing gene transfection to microinjection [1]. Fibroblasts secrete a non-rigid extracellular matrix that is rich in type I and/or type III collagen [2]. They are responsible for much of the synthesis of extracellular matrix in connective tissues and play major roles in wound healing. Human gallbladder fibroblasts (HGBF) are responsible for eicosanoid synthesis, specifically PGI_2. Chronic stimulation of HGBF results in up-regulated synthesis and release of PGI_2 which aids in formation of early acute cholecystitis [3]. HGBF are very versatile cells and could be used in studies of many conditions, such as the early stages of acute cholecystitis.

HGBF from ScienCell Research Laboratories are isolated from human gallbladder tissue. HGBF are cryopreserved at passage one and delivered frozen. Each vial contains $>5 \times 10^5$ cells in 1 ml volume. HGBF are characterized by their spindle morphology and immunofluorescence with antibody specific to fibronectin. HGBF are negative for HIV-1, HBV, HCV, mycoplasma, bacteria, yeast and fungi. HGBF are guaranteed to further expand for 15 population doublings at the condition provided by ScienCell Research Laboratories.

Recommended Medium

It is recommended to use Fibroblast Medium (FM, Cat. No. 2301) for the culturing of HGBF *in vitro*.

Product Use

HGBF are for research use only. It is not approved for human or animal use, or for application in *in vitro* diagnostic procedures.

Storage

Directly and immediately transfer cells from dry ice to liquid nitrogen upon receiving and keep the cells in liquid nitrogen until cell culture is needed for experiments.

Shipping

Dry ice.

Reference

[1] Conrad, G. W., Hart, G. W., Chen, Y. (1977) Differences in vitro between fibroblast-like cells from cornea, heart, and skin of embryonic chicks. J. Cell Sci. 26:119-137.

[2] Gabbiani, G., Rungger-Brandle, E., The fibroblast. In Tissue Repair and Regeneration (L. E. Glynn, ed.), pp 1-50. Handbook of Inflammation, Vol. 3. Amsterdam, Elsevier, 1981.
[3] Myers I. S., Evans T. C., Bartula L., Riva A., Kalley-Taylor B. (1995) Regulation of eicosanoid synthesis in fribroblasts from inflamed gallbladders. Mol. Cell Endocrinol. 115: 29-39

Instruction for culturing cells

Caution: Cryopreserved cells are very delicate. Thaw the vial in a 37^{o}C water bath and return them to culture as quickly as possible with minimal handling!

Set up culture after receiving the order:

1. Prepare a poly-$_{L}$-lysine coated flask (2 μg/cm^2, T-75 flask is recommended). Add 10 ml of sterile water to a T-75 flask and then add 15 μl of poly-$_{L}$-lysine stock solution (10 mg/ml, Cat. No. 0413). Leave the flask in incubator overnight (minimum one hour at 37^{o}C incubator).
2. Prepare complete medium: decontaminate the external surfaces of medium and medium supplements with 70% ethanol and transfer them to sterile field. Aseptically open each supplement tube and add them to the basal medium with a pipette. Rinse each tube with medium to recover the entire volume.
3. Rinse the poly-$_{L}$-lysine coated flask with sterile water twice and add 20 ml of complete medium to the flask. Leave the flask in the hood and go to thaw the cells.
4. Place the vial in a 37^{o}C water bath, hold and rotate the vial gently until the contents are completely thawed. Remove the vial from the water bath immediately, wipe it dry, sterilize the vial with 70% ethanol and transfer it to a sterile field. Remove the cap, being careful not to touch the interior threads with fingers. Using 1 ml eppendorf pipette gently re-suspend the contents of the vial.
5. Dispense the contents of the vial into the equilibrated, poly-$_{L}$-lysine coated culture vessels. A seeding density of 5,000 cells/cm^2 is recommended.
   Note: Dilution and centrifugation of cells after thawing are not recommended since these actions are more harmful to the cells than the effect of residual DMSO in the culture. It is also important that cells are plated in poly-$_{L}$-lysine coated flask that promotes cell attachment and growth.
6. Replace the cap or cover, and gently rock the vessel to distribute the cells evenly. Loosen cap if necessary to permit gas exchange.
7. Return the culture vessels to the incubator.

8. For best result, do not disturb the culture for at least 16 hours after the culture has been initiated. Change the growth medium the next day to remove the residual DMSO and unattached cells, then every other day thereafter.

Maintenance of Culture:

1. Change the medium to fresh supplemented medium the next morning after establishing a culture from cryopreserved cells.
2. Change the medium every three days thereafter, until the culture is approximately 70% confluent.
3. Once the culture reaches 70% confluence, change medium every other day until the culture is approximately 90% confluent.

Subculture:

1. Subculture the cells when they are over 90% confluent.
2. Prepare poly-$_L$-lysine coated flasks (2 μg/cm^2) one day before subculture.
3. Warm medium, trypsin/EDTA solution (T/E, ScienCell cat. no. 0103), trypsin neutralization solution (TNS, Cat. No. 0113), and DPBS to **room temperature**. We do not recommend warming the reagents and medium at 37^{o}C water bath prior to use.
4. Rinse the cells with DPBS.
5. Add 8 ml of DPBS first and then 2 ml of trypsin/EDTA solution into flask (in the case of T-75 flask); gently rock the flask to make sure cells are covered by trypsin/EDTA solution; incubate the flask at 37^{o}C incubator for 1 to 3 minutes or until cells are completely rounded up (monitored with inverted microscope). During incubation, prepare a 50 ml conical centrifuge tube with 5 ml of fetal bovine serum (FBS, Cat. No. 0500); transfer trypsin/EDTA solution from the flask to the 50 ml centrifuge tube (a few percent of cells may detached); continue incubate the flask at 37^{o}C for 1 minutes (no solution in the flask at this moment); at the end of trypsinization, one hand hold one side of flask and the other hand gently tap the other side of the flask to detach cells from attachment; check the flask under inverted microscope to make sure all cells are detached, add 5 ml of trypsin neutralization solution to the flask and transfer detached cells to the 50 ml centrifuge tube; add another 5 ml of TNS to harvest the residue cells and transfer it to the 50 ml centrifuge tube. Examine the flask under inverted microscope to make sure the cell harvesting is successful by looking at the number of cells left behind. There should be less than 5%.
   Note: Use ScienCell Research Laboratories' trypsin/EDTA solution that is optimized to minimize the killing of the cells by over trypsinization.

6. Centrifuge the 50 ml centrifuge tube (harvested cell suspension) at 1000 rpm for 5 min; re-suspend cells in growth medium.
7. Count cells and plate cells in a new, poly-$_{L}$-lysine coated flask with cell density as recommended.

Caution: Handling human derived products is potentially biohazardous. Although each cell strain tests negative for HIV, HBV and HCV DNA, diagnostic tests are not necessarily 100% accurate, therefore, proper precautions must be taken to avoid inadvertent exposure. Always wear gloves and safety glasses when working these materials. Never mouth pipette. We recommend following the universal procedures for handling products of human origin as the minimum precaution against contamination [1].

[1] Grizzle WE and Polt S. (1988) Guidelines to avoid personal contamination by infective agents in research laboratories that use human tissues. J Tissue Culture Methods. 11:191-9.