



Stem Cell Associated MicroRNA Array

Catalog Number AP-0007

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Introduction

Stem cells can differentiate into all of the specialized embryonic tissues during embryo development. In adult organisms, stem cells and progenitor cells act as a repair system for the body, replenishing specialized cells, in order to maintain the normal turnover of regenerative organs, such as blood, skin or intestinal tissues. Recent studies suggest that maintenance and differentiation of stem cells are regulated by microRNAs (miRNA). MiRNAs are small single-stranded RNA molecules (~20nt long), playing an important role in gene regulation and cell differentiation. MiRNAs are found to be differentially expressed in stem cells, such as brain-specific miR-124a and miR-9 molecules during neural lineage differentiation. In addition, miRNAs are also reported to regulate cancer stem cells. Signosis developed a miRNA array for targeting 69 miRNAs that are shown in literature associated with stem cell differentiation and maintenance. Profiling the expression of these miRNAs will help to reveal the functions of miRNAs in stem cells.

Based on its proprietary technology, Signosis has developed a highly sensitive and discriminative array technology for monitoring miRNA. It combines oligo-ligation assay-based detection and T7 transcription-based linear amplification, providing a highly sensitive and specific array assay. Via selection, ligation, and transcription of a pair of oligos that are hybridized to each specific miRNA, the targets are amplified and array monitored. By assigning unique tag sequences to individual isoforms, these isoforms can be easily differentiated. The whole procedure is simple and straightforward. We are currently offering an array that targets the 69 stem cells associated miRNAs, each with cited literature, which is able to facilitate the comparison and discovery of differentiated miRNA expression in different samples.

Principles of the technology

miRNAs are different from large messenger RNAs in three aspects; (1) miRNAs are small size molecules with quite a big difference in abundance, (2) mature miRNAs co-exist with their precursor pre-miRNA and pri-miRNA, differing only in length, and (3) many miRNAs are very closely related in sequences, such as isoforms, differing by only one or a few nucleotides. Therefore, the conventional microarray technologies cannot directly be applied to analyzing these molecules. A number of miRNA microarray products are commercially available, but they are either tedious in requiring pre-isolation of microRNA, lack discriminative power to differentiate isoforms, or are

not sensitive enough to monitor low abundant miRNAs.

In our array assay, one miRNA molecule is targeted by two oligos, each of which hybridizes a half molecule of the target miRNA to form a RNA/DNA duplex. When the sequences are perfectly matched, they are aligned with the miRNA and the joint can be ligated by DNA ligase (figure 1). A single nucleotide difference among miRNAs will block either the hybridization or the ligation, so that miRNA isoforms can be differentiated. Due to the small size of miRNA, the hybrid might not be stable; therefore we introduce the stacking sequences. By extending these two oligos along with their complementary oligos the stability is increased. Once the pair of oligos is ligated, the ligated molecules are subjected to linear amplification via T7 transcription into RNA in the presence of biotin-UTP, which are used as probes for array hybridization. To differentiate each isoform, we assigned unique tag sequences to the ligation oligos, so that single nucleotide differences are converted into unique tag sequences. Therefore, each isoform can be easily distinguished by array hybridization.

We offer the miRNA profiling assay kit to profile the expression of the 69 stem cell-associated miRNAs and their isoforms. The array can be used for human, mouse and rat samples. The procedure is simple and straight forward, including three steps: (1) mix the total RNA with provided oligos to form miRNA/oligo hybrids; (2) select the hybrids and remove free oligos, and ligate miRNA-directed pairing of oligos to become a single DNA; and (3) amplify the ligated DNA with T7 transcription.

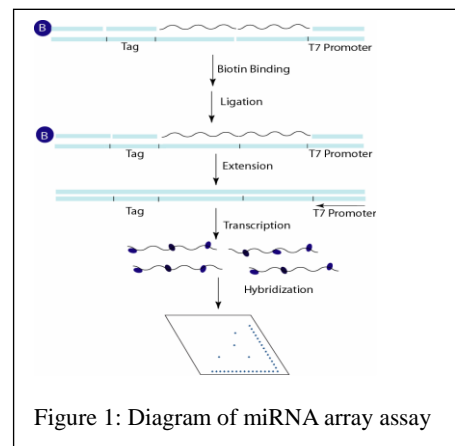


Figure 1: Diagram of miRNA array assay

Materials provided with the kit

15 µl Stem Cell Oligo Mix (-20°C)
5 µl Array Detection Oligo
400 µl Annealing buffer (RT)
15 µl Magnetic streptavidin beads (4 °C)
120 µl Beads binding buffer (RT)
1.0 ml Bead wash buffer (RT)
6 µl Ligase (-20°C)
250 µl Ligation buffer (-20°C)
60 µl Extension mix (-20°C)
60 µl Labeling mix (-20°C)
6 µl T7 RNA polymerase (-20°C)
30ml 1x Hybridization buffer (RT)
30ml 5x Hybridization wash buffer (RT)
60ml Blocking buffer (RT)
50 µl Streptavidin-HRP conjugate (4 °C)
40ml 5x Detection wash buffer (RT)
1.8ml Substrate A (4 °C)
1.8ml Substrate B (4 °C)
3 Stem cell array membranes (RT)
3 Detection sheets (RT)

Material required but not provided

Magnetic stand (96 well plate)
PCR machine
Hybridization oven
Washing tray
50ml Centrifuge tubes (Corning tubes are recommended, cat#430290) or hybridization bottles
0.2ml PCR tube
Alpha Innotech image or equivalent image system or X-ray film
RNase free water

Reagent preparation before starting experiment

- Dilute the 5x Hybridization washing buffer and 5x Detection washing buffer to 1x buffer
1x Hybridization washing buffer:
30ml 5x Hybridization washing buffer
120ml ddH₂O
1x Detection washing buffer:
40ml 5x Detection washing buffer
160 ml ddH₂O
- Prewarm 1x Hybridization buffer, 1x Hybridization wash buffer at 42°C for 1 hour or until the buffers are clear without visible precipitation before using.
- Pre-hybridization can be done during T7 RNA transcription at the Step 4.

Assay Procedure

Assay Procedure

1. Annealing of miRNA with Oligo mix

- (1) Sample preparation
X µl 1-5ug total RNA or 10ng isolated miRNA
5 µl Stem cell oligo mix
1 µl Array Detection Oligo
20 µl Annealing buffer
X µl ddH₂O

40ul

- (2) Incubate on a PCR machine at 72°C for 5 minutes and 53°C for 90 minutes.

2. Selection of miRNA/oligo hybrids

- (1) Beads washing
- Resuspend the beads by gently tapping the tube to obtain a homogeneous suspension.
 - Transfer 5 µl beads to a 0.2ml PCR tube (the size of the tube that should fit into the magnetic stand.
 - Add 100 µl of annealing buffer to the tube, place onto the magnetic stand for 30 seconds.
 - Aspirate out the liquid.
 - Remove the tube from magnetic stand.

- (2) Beads selection
- Add 40 µl of Bead binding buffer to 40 µl annealed miRNA/oligo hybrid mix from Step 1, transfer to the washed beads and resuspend the beads in the solution.
 - Incubate at 37°C for 30 minutes.
 - Place the bead mixture on the magnetic stand for 30 seconds, and aspirate out the buffer. The beads will remain on the side of tube.
 - Remove the tube from the magnetic stand and add 100 µl of Bead wash buffer to resuspend the beads, pipette gently up and down, and place the tube on the magnetic stand for 30 seconds, aspirate the buffer.
 - Repeat the washing step once.

3. Ligation of miRNA-directed oligos to form a single molecule

- (1) Add 50 µl of Ligation buffer to resuspend the beads, pipette gently up and down, place the tube on the magnetic stand for 30 seconds, and aspirate the buffer.
(2) Remove the tube from the magnetic stand. Add 20 µl of ligation buffer to resuspend the beads, then add 2µl ligase to the resuspended beads and incubate at 37°C for 90 minutes.

4. T7 RNA transcription of ligated molecule

- (1) Add 100 μ l Bead washing buffer directly to 20 μ l ligation reaction mix from Step 3, place the tube on the magnetic stand for 30 seconds, and aspirate the buffer.
- (2) Remove the tube from the magnetic stand and add 20 μ l of Extension mix to resuspend the beads.
- (3) Incubate the mixture on PCR machine at 94°C for 2 minutes, 54°C for 1 minute, 72°C for 1.5 minute, and 94°C for 30 seconds.
- (4) Place the reaction tube on the magnetic stand for 10 seconds. **Immediately** transfer the 20 μ l of the extension mix to a fresh tube (keep the solution and toss the beads).
- (5) Add 20 μ l of Labeling mix and 1 μ l of T7 RNA polymerase to the tube.
- (6) Incubate the mixture at 37°C for 1 hour.
- (7) The transcribed RNA is ready for hybridization.

5. Pre-hybridization and hybridization

- (1) Place each array membrane into a 50 ml tube. Wet the membrane by filling the tube with dH₂O, then carefully decant the water. The side of the membrane with the spotted oligos should face into the middle of the tube.
- (2) Add 4 ml of prewarmed 1x Hybridization buffer to each tube. Incubate the tubes in a hybridization oven at 42°C for at least 30-60 minutes.
- (3) Decant the hybridization buffer and replace with 4ml of prewarmed 1x Hybridization buffer. Add 40 μ l of transcribed RNA to prehybridized membrane and incubate overnight in a hybridization oven at 42°C.
- (4) Decant the hybridization mixture from each tube and wash each membrane as follows:
 - Rinse the membrane with 20 ml Hybridization washing buffer, and decant liquid.
 - Incubate the membrane with 20 ml Hybridization wash buffer at 42°C for 20 minutes. Decant liquid.

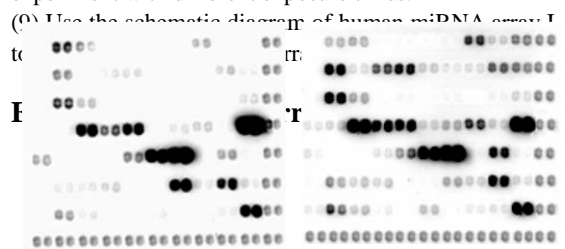
6. Detection

- (1) Using forceps, carefully transfer the membrane from the hybridization tube to a container (an empty 200 μ l pipette tips box). Each box could have two membranes, one at each side of the box.
- (2) Rinse the membrane with 10 ml of 1X Detection wash buffer.
- (3) Block the membrane with 15 ml of Blocking buffer for 30 minutes at room temperature with moderate shaking.
- (4) Dilute 15 μ l of Streptavidin-HRP conjugate with 1 ml of the 1X Blocking buffer and transfer to the container. **Do not** add HRP diluted solution directly onto the membrane.
- (5) Continue shaking the membrane for 45 min at room temperature.
- (6) Decant the Blocking buffer and wash three times at room temperature with 15 ml of 1x Detection washing buffer, 10 minutes each wash.

(7) Mix equal amounts of Substrate A and B. Place the membrane on the bottom side of detection sheet on a flat surface and overlay the membrane with 1 ml of substrate solution. To ensure that the solution remains evenly distributed over the membrane when enveloped by the detection sheet: gently lower the top side of the detection sheet halfway over the membrane then pull back up slightly to allow the solution to flow back over the membrane. Then slowly lay the top sheet down completely without trapping air bubbles. Incubate at room temperature for 5 minutes.

(7) Mix equal amounts of Substrate A and B. Place the membrane on the bottom side of detection sheet on a flat surface and overlay the membrane with 1 ml of substrate solution. To ensure that the solution remains evenly distributed over the membrane when enveloped by the detection sheet: gently lower the top side of the detection sheet halfway over the membrane then pull back up slightly to allow the solution to flow back over the membrane. Then slowly lay the top sheet down completely without trapping air bubbles. Incubate at room temperature for 5 minutes.

(8) Remove excess substrate by gently applying pressure over the top sheet using a paper towel. Expose the membranes using either Hyperfilm ECL (2-10 min) or a chemiluminescence imaging system (i.e., FluorChem imager from Alpha Innotech). With either method, experiment with different exposure times.



2 μ g total RNA from mouse and mouse embryo was used for miRNA array assay respectively, and the arrays were detected with a chemiluminescence imaging system.

Trouble Shooting

Signals are too weak

- Total RNA may not contain small RNA
- RNA may be degraded
- If signals of the alignment spots are weak as well, the incubation of the membrane with Streptavidin HRP conjugate may be too short or the exposure time may be too short.

Uneven background

- Substrate was not evenly overlaid on the membrane

References

References

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Schematic diagram of stem cell-associated miRNA array

Let-7a	Let-7b	Let-7c	Let-7d	Let-7e	Let-7f	miR-1	miR-9	miR-10a	miR-15a
miR-15b	miR-16	miR-17-5p	miR-19a	miR-19b	miR-21	miR-25	miR-34a	miR-99a	miR-122a
miR-124a	miR-125a	miR-125b	miR-126	miR-133a	miR-133b	miR-146a	miR-146b	miR-155	miR-181a
miR-181b	miR-181c	miR-199a	miR-199b	miR-9-1	miR-10b	miR-22	miR-23a	miR-24	miR-26a
miR-26b	miR-27a	miR-27b	miR-30a-3p	miR-30a-5p	miR-30b	miR-30c	miR-92b	miR-93	miR-101-1
miR-103	miR-106a	miR-106b	miR-107	miR-128a	miR-128b	miR-132	miR-134	miR-135b	miR-141
miR-149	miR-150	miR-181d	miR-218	miR-296	miR-213	miR-96	miR-193a	miR-193b	U6