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## Inflammation Luciferase Reporter Vector Set

Catalog Number LR-3005

(For Research Use Only)

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### Introduction

Inflammation is a complex physiological reaction to noxious stimuli including tissue stress, injury and infection. Dysregulation of inflammation often occurs in people and can cause a variety of pathologies, ranging from chronic inflammation, to autoimmunity, to cancer. What leads to the inducible expression of inflammatory genes at the level of gene transcription is a crucial question, and the answer lies in transcription factors (TF). TF-based reporters are often used to monitor the activation of signaling pathways.

Signosis has developed a set of luciferase reporter vectors (NFkB, AP1, IRF, ATF3, Stat3, C/EBP, RUNX1, and Negative control) to facilitate inflammation study.

### Recommend transfection and assay

We recommend using FuGENE 6 (Roche) for the transfection of pTF-Luc reporter vectors. For difficult-to-transfect cell type such as primary cells, we recommend using Fugene HD (Roche) for the transfection. The transfection can be done in 6-well or 12 well plates.

The following protocol is designed for adherent cultures in **6-well** plates using FuGENE 6. If you use a different size of plate or flasks, adjust the components in proportion to the surface area of your container.

Below is the assay procedure for each of the 8 vectors:

1. For each of the 8 vectors, plate  $1-4 \times 10^5$  cells in 2 ml of growth medium containing serum without antibiotics in a 6-well culture plate at one day before transfection, which will yield 50-80% confluence on the day of transfection.

2. dilute 0.5-1  $\mu\text{g}$  of the reporter vector with 100  $\mu\text{l}$  of serum-free culture medium, and in a separate tube, dilute 3  $\mu\text{l}$  FuGENE 6 Reagent with 100  $\mu\text{l}$  of serum-free culture medium (add transfection reagent directly into the medium and don't touch the wall of the tube). Add the diluted reporter vector to the diluted transfection reagent and gently mix. Incubate for 15-30 min at room temperature. Once the FuGENE 6 Reagent is diluted, it needs to use within 45 min.

3. Add 200  $\mu\text{l}$  of DNA/FuGENE complex to on the cells in a drop-wise manner. Evenly distribute the complex by gently rocking the plate back and forth. Incubate the cells at  $37^\circ\text{C}$  in a CO<sub>2</sub> incubator for overnight.

4. If the starvation is required, replace the medium with serum free or low serum medium (0.2% serum) for 6 -16 hours, and treat the cell with the selected stimulus for 8-14 hours.

5. Alternatively, to study the effects of a gene of interest, cotransfect each pTF-Luc with a gene expression vector of interest.

6. Lyse the attached cells by adding lysis buffer (Promega, Luciferase Assay System) to each well. Use approximately 200  $\mu\text{l}$  per well for a 6-well plate. To detach cells from the plate, freeze and thaw the plate once and pipette the mixture up and down. Transfer the cell lysate/buffer solution to a clean 1.5-ml microcentrifuge tube, which is ready for luciferase assay or store at  $-80^\circ\text{C}$  for the future use. Assay for luciferase activity following the instructions given by the supplier (Promega, Luciferase Assay System, cat.# E1500).

### E. coli transform to propagate the plasmids

1. Transform *E. coli* competent cells with 1 $\mu\text{l}$  plasmid provided.

2. Plate the transformed cells on LB plates containing 100  $\mu\text{g}/\text{mL}$  Ampicillin and grow overnight at  $37^\circ\text{C}$ .

3. Transfer a single colony to 1-2 ml LB medium containing 100  $\mu\text{g}/\text{mL}$  and shake at  $37^\circ\text{C}$  overnight.

4. Prepare plasmids and check on gel.