

Mouse MIP-1α ELISA Catalog Number EA-2409

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Introduction

Macrophage Inflammatory Protein-1 (MIP-1 α) is a member of the C-C subfamily of chemokines that exhibit a variety of proinflammatory activities in vitro including leukocyte chemotaxis. MIP-1 is a major factor produced by macrophages after stimulated with bacterial endotoxins. It activates granulocytes (neutrophils, eosinophils and basophils) which can lead to acute neutrophilic inflammation. It also induces the synthesis and release of other pro-inflammatory cytokines such as IL-1, IL-6 and TNF α from fibroblasts and macrophages. In addition to its proinflammatory activities, MIP-1 α inhibits the proliferation of hematopoietic stem cells in vitro and in vivo.

Principle of the assay

MIP-1a ELISA is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes rabbit anti-mouse MIP-1a a antibodies for immobilization on the microtiter wells and rabbit anti-mouse MIP-1a antibodies along with streptavidin conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two antibodies, resulting in the MIP-1 α molecules being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentration of MIP-1 α is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.

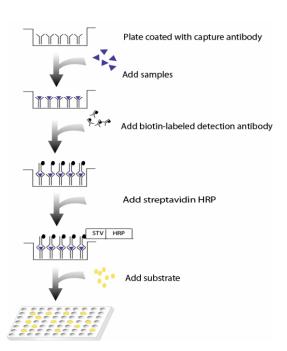


Diagram of ELISA

Materials provided with the kit

- 96 well microplate coated with rabbit anti-mouse MIP-1α antibodies (4°C).
- Biotin labeled rabbit anti-mouse MIP-1 α antibodies (-20°C).
- Streptavidin-HRP conjugate (4°C).
- Recombinant mouse MIP-1α standard (1000ng/ml) (-20°C).
- 1X Diluent buffer (4°C).
- 5X Assay wash buffer (RT)
- Substrate (4°C).
- Stop Solution (4°C).

Material required but not provided

- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

(For Research Use Only)

Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer 40ml 5x Assay wash buffer 160ml ddH2O
- Dilute 500 times of mouse recombinant MIP-1α (1000ng/ml) with 1X Diluent buffer to 2000pg/ml and then 2-fold serial dilutions.
- Dilute 400 times of biotin labeled rabbit anti-mouse MIP-1α antibodies with 1X Diluent buffer before use.
- Dilute 200 times of streptavidin-HRP with 1X Diluent buffer before use.

Assay procedure

1. Cut the sealing film over the plate and remove it from the desired number of well strips. Make sure the rest of wells are well sealed.

2. Add $100 \square \mu l$ of Standard, control, or sample per well and incubate for 1 hour at room temperature with gentle shaking.

3. Aspirate each well and wash by adding 200μ l of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Complete removal of liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.

4. Add 100µl of diluted biotin-labeled rabbit anti-mouse MIP-1 α antibodies to each well and incubate for 1 hour at room temperature with gentle shaking.

5. Repeat the aspiration/wash as in step 3.

6. Add 100 μ l of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.

7. Repeat the aspiration/wash as in step 3.

8. Add 100 μl substrate to each well and incubate for 5-30 minutes.

9. Add $50\mu l$ of Stop solution to each well. The color in the wells should change from blue to yellow.

10. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.

Example of standard curve

