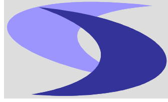


Signosis
Innovative Plate Assay Solutions

Mouse IL-12 ELISA

Catalog Number EA-2514 **(For Research Use Only)**

Introduction

Interleukin 12 (IL-12) is an interleukin that plays important roles in the activities of natural killer cells and T lymphocytes, the differentiation of naive T cells into Th1 cells, the growth and function of T cells, the production of interferon-gamma (IFN-γ) and tumor necrosis factor-alpha (TNF-α) from T and natural killer (NK) cells, and IL-4 mediated suppression of IFN-γ. It also plays a role in anti-angiogenic activity by increasing production of IFN-γ, which in turn increases the production of IP-10. IL-12 is naturally produced by dendritic cells, macrophages and mouse B-lymphoblastoid cells in response to antigenic stimulation.

Principle of the assay

IL-12 ELISA is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes goat anti-mouse IL-12 for immobilization on the microtiter wells and biotinated goat anti-mouse IL-12 antibodies along with streptavidin conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two antibodies, resulting in the IL-12 molecules being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentration of IL-12 is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.

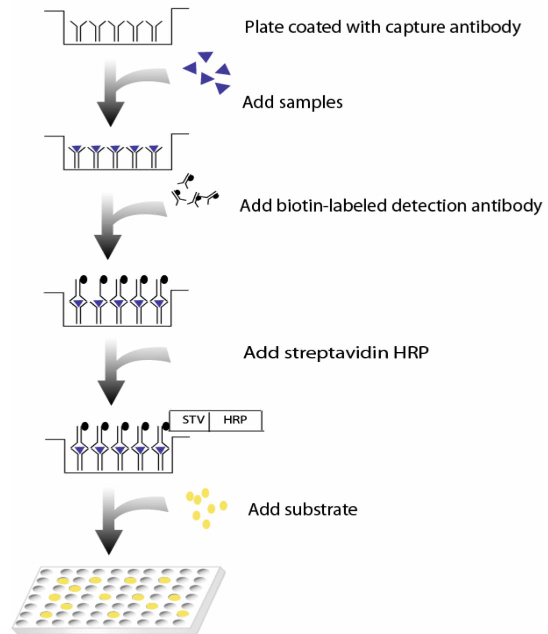


Diagram of ELISA

Materials provided with the kit

- 8x12 96-well microplate coated with goat anti-mouse IL-12 antibodies (4°C)
- Biotin labeled goat anti-mouse IL-12 antibodies (-20°C)
- Streptavidin-HRP conjugate (4°C)
- Recombinant mouse IL-12 standard (-20°C)
- 1X Diluent buffer (4°C)
- 5X Assay wash buffer (4°C)
- Substrate (4°C)
- Stop Solution (4°C)

Material required but not provided

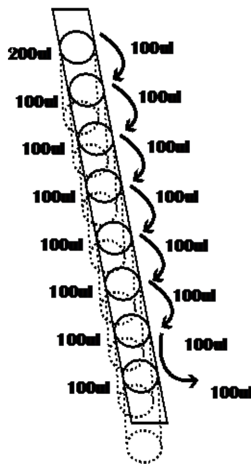
- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer
  40ml 5x Assay wash buffer
  160ml ddH2O
- Use serum-free conditioned media or original or 10-fold diluted sera. Sera can be diluted with 1 X Diluent buffer. When serum-containing conditioned media is required, be sure to use serum as a control.
- Dilute 50 times of mouse recombinant IL-12 (200ng/ml) with 1X Diluent buffer to 4000pg/ml and then 2-fold serial dilutions. Add 4ul Mouse Recombinant IL-12 in 200ul 1X Diluent Buffer (See Step 2 below for detailed instruction)
- Dilute 400 times of biotin labeled goat anti-mouse IL-12 antibodies with 1X Diluent buffer before use.
- Dilute 200 times of streptavidin-HRP with 1X Diluent buffer before use.

Assay procedure

1. Calculate the number of samples to decide how many strips need to be used.
2. Add 100 μl of Standard, control, or sample per well and incubate for 1 hour at room temperature with gentle shaking.



a. Add 200ul 1X Diluent buffer to the 1st well. Add 100ul 1X Diluent Buffer to the rest wells of strip.
b. Add appropriate amount of protein recombinant (follow instruction in "Reagent Preparation")
c. Mix dilutions in 1st well and transfer 100ul from the 1st well to the next dilution. (See picture) Incubate each well for 1 hr at room temperature with gentle shaking

3. Aspirate each well and wash by adding 200 μl of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Complete removal of liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.
4. Add 100 μl of diluted biotin-labeled goat anti-mouse IL-12 antibodies to each well and incubate for 1 hour at room temperature with gentle shaking.
5. Repeat the aspiration/wash as in step 3.
6. Add 100 μl of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.
7. Repeat the aspiration/wash as in step 3.
8. Add 100 μl of substrate to each well and incubate for 5-30 minutes.
9. Add 50 μl of Stop solution to each well. The color in the wells should change from blue to yellow.
10. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.