



Mouse IL-17A ELISA

Catalog Number EA-2516

(For Research Use Only)

Introduction

Interleukin-17a (IL-17a) is a cytokine primarily produced by activated T cells to regulate local tissue inflammation. IL-17A can induce inflammatory cytokine production through the regulation of NF κ B and MAPK family pathways. Elevated levels of IL-17 are associated with several chronic inflammatory diseases, including asthma, rheumatoid arthritis, and multiple sclerosis and has become an important potential target for their treatment. Understanding the conditions that alter the expression of this vital cellular messenger is important for unraveling the mechanisms of these and other diseases and for developing therapeutics.

Principle of the assay

IL-17A ELISA is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes goat anti-mouse IL-17A for immobilization on the microtiter wells and biotinylated goat anti-mouse IL-17A antibodies along with streptavidin conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two antibodies, resulting in the IL-17A molecules being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentration of IL-17A is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.

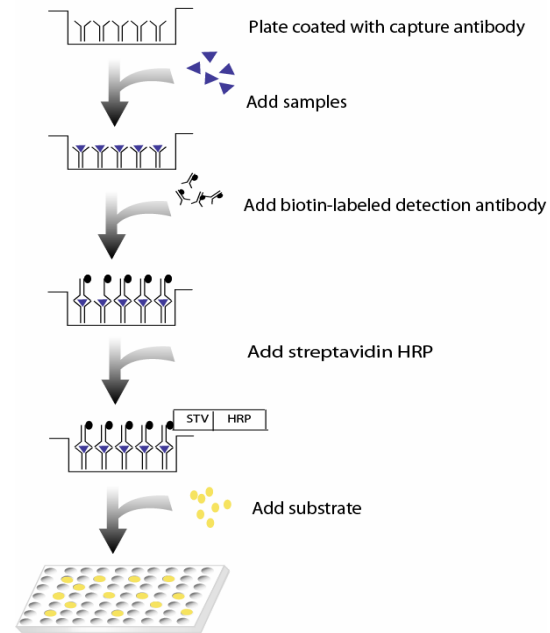


Diagram of ELISA

Materials provided with the kit

- 8x12 96-well microplate coated with goat anti-mouse IL-17A antibodies (4°C)
- Biotin labeled goat anti-mouse IL-17A antibodies (-20°C)
- Streptavidin-HRP conjugate (4°C)
- Recombinant mouse IL-17A standard (-20°C)
- 1X Diluent buffer (4°C)
- 5X Assay wash buffer (4°C)
- Substrate (4°C)
- Stop Solution (4°C)

Material required but not provided

- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

Reagent preparation before starting experiment

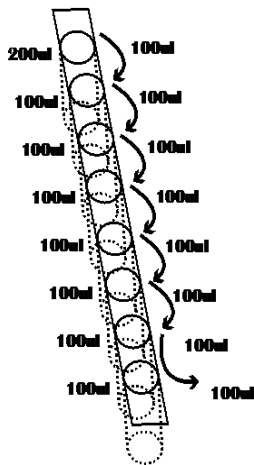
- Dilute the 5x Assay wash buffer to 1x buffer
40ml 5x Assay wash buffer
160ml ddH₂O

Use serum-free conditioned media or original or 10-fold diluted sera. Sera can be diluted with 1 X Diluent buffer. When serum-containing conditioned media is required, be sure to use serum as a control.

- Dilute 50 times of mouse recombinant IL-17a (200ng/ml) with 1X Diluent buffer to 4000pg/ml and then 2-fold serial dilutions. Add 4ul Mouse Recombinant IL-17a in 200ul 1X Diluent Buffer (See Step 2 below for detailed instruction)
- Dilute biotin labeled rabbit anti-mouse IL-17a antibodies 1:400 with 1X Diluent buffer before use.
- Dilute streptavidin-HRP 1:200 with 1X Diluent buffer before use.

Assay procedure

1. Calculate the number of samples to decide how many strips need to be used.
2. Prepare standard according to diagram.



1. Add 200 μ l 1X Diluent buffer to the 1st well. Add 100 μ l 1x Diluent Buffer to the rest of the wells in the strip.
2. Add 4 μ l of standard to the first well.
3. Mix dilution in 1st well and transfer 100 μ l from the first well to the 2nd well.
4. Repeat mix and transfer 100 μ l into each additional well as pictured.

3. Add 100 μ l of sample per well and incubate for 1 hour at room temperature with gentle shaking.
4. Aspirate each well and wash by adding 200 μ l of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Completely remove liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.
5. Add 100 μ l of diluted biotin-labeled rabbit anti-mouse IL-17a antibody to each well and incubate for 1 hour at room temperature with gentle shaking.
6. Repeat the aspiration/wash as in step 4.

7. . Add 100 μ l of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.
8. Repeat the aspiration/wash as in step 4.
9. Add 100 μ l of substrate to each well and incubate for 10-30 minutes.
10. Add 50 μ l of Stop solution to each well. The color in the wells should change from blue to yellow.
11. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.