

## **Mouse IL-23 ELISA**

**Catalog Number EA-2517** 

### Introduction

The cytokine Interleukin-23 (IL-23) is a heterodimer comprised of a p19 Interleukin-23 subunit and a p40 subunit of IL-12. The complete IL-23 is produced in activated macrophages and dendritic cells. IL-23 acts on memory T cells to induce proliferation and IFNy and IL-17 production. IL-23 is known to play a role in multiple sclerosis, inflammatory bowel disease, and psoriasis. In fact, a monoclonal antibody that recognizes IL-23 and IL-12 is approved to treat psoriasis and psoriatic arthritis.

#### Principle of the assay

IL-23 ELISA is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes rabbit anti-mouse IL-23 for immobilization on the microtiter wells and biotinated rabbit anti-mouse IL-23 antibodies along with streptavidin conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two antibodies, resulting in the IL-23 molecules being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentration of IL-23 is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.

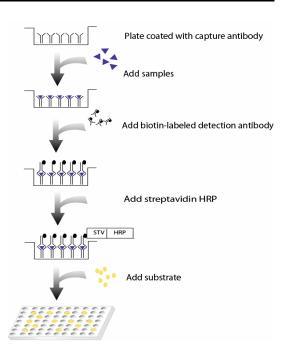


Diagram of ELISA

#### Materials provided with the kit

- 8x12 96-well microplate coated with rabbit anti-mouse IL-23 antibodies (4°C).
- Biotin labeled rabbit anti-mouse IL-23 antibodies (-20°C).
- Streptavidin-HRP conjugate (4°C).
- Recombinant mouse IL-23 standard (1000ng/ml) (-20°C).
- 1X Diluent buffer (4°C).
- 5X Assay wash buffer (RT)
- Substrate (4°C).
- Stop Solution  $(4^{\circ}C)$ .

#### Material required but not provided

- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

#### (For Research Use Only)

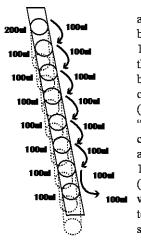
# Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer 40ml 5x Assay wash buffer 160ml ddH2O
- Use serum-free conditioned media or original or 10fold diluted sera. Sera can be diluted with 1 X Diluent buffer. When serum-containing conditioned media is required, be sure to use serum as a control.
- Dilute 200 times of mouse recombinant IL-23 (1000ng/ml) with 1X Diluent buffer to 5000pg/ml and then 2-fold serial dilutions.
- Dilute 400 times of biotin labeled rabbit anti-mouse IL-23 antibodies with 1X Diluent buffer before use.
- Dilute 200 times of streptavidin-HRP with 1X Diluent buffer before use.

#### Assay procedure

1. Cut the sealing film over the plate and remove it from the desired number of wells. Make sure the rest of wells are well sealed.

2. Add 100  $\mu$ l of Standard, control, or sample per well and incubate for 1 hour at room temperature with gentle shaking.



a. Add 200ul 1X Diluent buffer to the 1<sup>st</sup> well. Add 100ul 1X Diluent Buffer to the rest wells of strip.
b. Add appropriate amount of protein recombinant (follow instruction in "Reagent Preparation")
c. Mix dilutions in 1<sup>st</sup> well and transfer 100ul from the 1<sup>st</sup> well to the next dilution. (See picture) Incubate each well for 1 hr at room temperature with gentle shaking

3. Aspirate each well and wash by adding  $200\,\mu$ l of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Complete removal of liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.

4. Add 100  $\mu$ l of diluted biotin-labeled rabbit anti-mouse IL-23 antibodies to each well and incubate for 1 hour at room temperature with gentle shaking.

5. Repeat the aspiration/wash as in step 3.

6. Add 100  $\mu$ l of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.

7. Repeat the aspiration/wash as in step 3.

8. Add 100  $\mu l$  of substrate to each well and incubate for 5-30 minutes.

9. Add  $50\,\mu$ l of Stop solution to each well. The color in the wells should change from blue to yellow.

10. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.