

Beta Actin ELISA Kit

Catalog Number EA-6001

(For Research Use Only)

Introduction

Beta-Actin is highly conserved proteins and is ubiquitously expressed in all eukaryotic cells at a constant level regardless of experimental treatment or technical procedure in most cases. Therefore, measurement of beta-Actin is generally used as an internal control for experimental error. Signosis has provided b-actin sandwich ELISA assay specifically to detect the endogenous levels of beta-actin, which can be used for internal control for most of ELISA assays with human, mouse and rat samples.

Principle of the assay

Beta-actin ELISA is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes a rabbit polyclonal beta-actin capture antibody for immobilization on the microtiter wells, and a mouse monoclonal beta-actin detection antibody along with antimouse conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two antibodies, resulting in the beta-actin molecules being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of stop solution changing the color to yellow. The concentration of beta-actin is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.

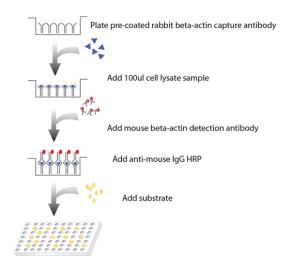


Diagram of ELISA

Materials provided with the kit

- 12x8 strip microplate coated with beta-actin capture antibody (4°C).
- Mouse beta-actin detection antibody (4°C).
- Anti-mouse IgG HRP (-20°C)
- 1X Diluent buffer (4°C)
- 5X Assay wash buffer (RT)
- Substrate (4°C)
- Stop Solution (4°C)

Material required but not provided

- Cell lysis buffer for ELISA (EA-0001)
- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer 40ml 5x Assay wash buffer 160ml ddH2O
- Sample preparation: Add 200ul 1X cell lysis buffer to one well of 12-well plate and pipetting up and down to detach the cells and incubate on ice for 5 minutes. Transfer the cell to 1.5ml tube and sonicate briefly on ice. The cell lysate was centrifuged for 5 minutes at 12000rpm. The supernatant is transferred to a new tube for downstream assay. If your sample is a well of 96-well plate (10⁴ cells), you can add 100ul 1X Cell lysate buffer in the well and directly use the cell lysate for the assay.
- Dilute 400 times of mouse monoclonal beta-actin antibody with 1X Diluent buffer before use.
- Dilute 1000 times of anti-mouse HRP conjugate with 1X Diluent buffer before use.

Assay procedure

- 1. Cut the sealing film over the plate and remove it from the needed number of well strips. Make sure the rest of wells are well sealed.
- 2. Add 100 μ l of sample per well and incubate at 37°C for two hours.
- 3. Aspirate each well and wash by adding $200\mu l$ of 1X Assay wash buffer. Repeat the process two times for a total of three washes. Complete removal of liquid at each wash by inverting the plate against a pile of clean paper towels.
- 4. Add $100\mu l$ of diluted mouse monclonal beta-actin detection antibody to each well and incubate for one hour at room temperature with gentle shaking.
- 5. Repeat the aspiration/wash as in step 3.
- 6. Add 100 µl of diluted anti-mouse IgG HRP conjugate to each well and incubate for one hour at room temperature.
- 7. Repeat the aspiration/wash as in step 3.
- 8. Add $100\mu l$ of substrate to each well and incubate for 15-30 minutes.
- 9. Add 50µl of Stop solution to each well. The color in the wells should change from blue to yellow.
- 10. Determine the optical density of each well with a microplate reader at 450 nm within 45 minutes.

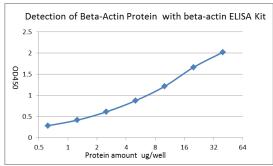


Figure 1. Detection of beta-actin protein with beta-actin ELISA kit. HeLa cells were grown in a 12-well plate. Cell lysate was prepared with Cell Lysis Buffer, 1:2 serially diluted with 1 x Diluent buffer and subject to ELISA assay.